

## Power On

1. Power on main power
2. Power on Source and Detector
3. Power on PC and monitor

## System Initialization

1. Log in to PC and software
2. Turn on UV lamp (if planning on doing a run)
  - a. Select Tools | Lamp Control | Turn Lamp On
  - b. Allow 20-30 minutes for equilibration
3. Initialize Camera and Temperature
  - a. Select Tools | Initialize Camera
  - b. Select Tools | Read CCD Temperature (wait for 5 minutes to reach CCD temperature)
  - c. Temperature must be between -25°C to -35°C

## File Set-Up

1. Creating a new file
  - a. Category Structure
    - i. Organize by product, user, month, etc.
  - b. File Structure
    - i. Name by product, date, user, etc.
2. Opening a File
  - a. Open a file for date review
  - b. Open a previously stored method
    - i. When opening a method make sure to rename the file immediately
3. Saving a File or Method
  - a. Save a File at the conclusion of the run
    - i. Enter the password to save File
  - b. Save a Method

Methods can be saved with method parameters and standard and blank data (if planning on performing a test be sure to rename the method as a file)

## Software Overview

1. Views (Quick Buttons and View Menu)
  - a. Report View
    - i. Method Parameters
    - ii. Standard Information
    - iii. Absorbance Values
    - iv. % Dissolved Results

- b. Spectral View
  - i. Wavelength Calibration File
  - ii. Blank Spectra
    - 1. Intensity
  - iii. Standard Spectra
    - 1. Absorbance
  - iv. Sample Spectra
    - 1. Absorbance
  - v. Spectra List
    - 1. List of all acquired and pending spectra
- c. Dissolution Curve View
  - i. Real-time plotting of dissolution results
- d. Image View
  - i. Color representation of intensity (Transmittance)
- e. Event Log (View Menu only)
  - i. Audit Trail to track method changes

## Method Parameter Set-Up

### 1. *General*

- a. Noted, Media Type informational
- b. Volume critical to calculation
- c. RPM critical if in bath control

### 2. *Sample*

- a. Unit Dose (mg) for Ingredient A and B critical to calculation
- b. Sample ID, Study Number, Storage Conditions informational

### 3. *Run*

- a. Exposure Time
  - i. Exposure Auto Set
    - 1. Place probes in blank media
    - 2. Select Exposure Auto Set
      - a. System will systematically find optimal exposure time
      - b. Optimal exposure time will populate exposure time field in method parameters
- b. Pathlength (Informational Only)
- c. Number of Scans (Signal Averaging)
  - i. Scope
    - 1. Used to improve signal to noise ratio
    - 2. Will provide better representation of true concentration in vessel
  - ii. Setting
    - 1. Traditional Application 3 to 4 scans
    - 2. Low Absorbing Application 10 plus scans
  - iii. Precaution
    - 1. Selecting too many scans can freeze the computer if sampling interval is not long enough
    - 2. Rule of Thumb: Assume 2 - 3 seconds for every scan due to the need to collect background and process data
- d. Phases (Timepoints)

- i. Divide dissolution curve into relevant segments
  1. Disintegration or Capsule Burst Section
  2. Rate of Release section
  3. Plateau section
- ii. Select interval relevant to requirement for characterization
  1. Early Timepoints
    - a. 10 or 15 second interval for first 5 - 15 minutes
      - i. More variability
      - ii. Better define formulation
  2. Middle Timepoints
    - a. 30 seconds to 1 minute for next 15 - 45 minutes
      - i. Characterize rate of release
      - ii. Identify potential production issues
        1. Content Uniformity
        2. Blending
        3. Compaction Pressure
        4. Etc.
  3. End Timepoints
    - a. 1 minute to 15 minutes for remainder of test

#### 4. Analytical

- a. Calculation Type
  - i. Single Component Analysis
    1. Analytical Wavelength (Single)
      - a. Defines calculation wavelength
      - b. Able to change before, during, and after run
  - ii. Multi-Component Analysis
    1. Wavelength Region (Multi)
      - a. Defines the portion of the spectrum used for data analysis
      - b. Region should encompass spectral regions that include absorbance peaks for both analytes A and B
      - c. Able to change before, during, and after run
- b. Baseline Correction (Single Component Analysis)
  - i. Scope
    1. Accounts for UNDISSOLVED PARTICULATES in solution (scattering effects)
  - ii. Types
    1. Single Wavelength
      - a. Reference Wavelength
    2. Double Wavelength
      - a. Two Point Perpendicular Drop
    3. Average Over Range
      - a. Define baseline by Range
    4. First Derivative
      - a. Inflection points at peak maxima locations of original spectrum
      - b. Analytical wavelength should be set to the largest positive or negative peak
      - c. Used when there is no flat portion of spectrum available for reference wavelength
    5. Second Derivative

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- a. Slope of the Slope
      - b. Attempting to decipher the effect of an active peak from a placebo
      - c. Poorly understood by regulatory
    - c. Component Selection (Multi-Component Analysis)
      - i. Select Ingredient A for Component 1
      - ii. Select Ingredient B for Component 2
      - iii. Select Baseline Shift Type(s):
        - 1. Constant
          - a. Spectrum shifts either up or down by a constant amount across UV spectrum
        - 2. Linear
          - a. Spectrum shifts by an amount that varies in linear manner with respect to wavelength
        - 3. Light Scattering
          - a. Curved baseline shift by Rayleigh scattering
        - 4. Custom
          - a. Ad-hoc image of excipient/placebo solution
    - d. Standard Mixtures (Multi-Component Analysis)
      - i. Scope
        - 1. Accounts for interactions between both ingredients
      - ii. Select Ad-hoc images for mixtures
      - iii. Input ingredient concentrations of the selected mixture
- 5. Report**
- a. Customize report format
  - b. Customize format of displayed values (decimal points)
  - c. Select channels of interest
    - i. Running two different formulations
    - ii. Running two different strengths
    - iii. Running two different medias
  - d. Multi-Component
    - i. Select which ingredient or both to display
    - ii. Average Fit Error option
      - 1. To ensure proper wavelength region and baseline shift options that results with the lowest Average Fit Error
    - iii. Display Observed vs. Calculated Plot option
      - 1. Select specific time point during dissolution run
      - 2. To ensure proper wavelength region and baseline shift options that results with Calculated Spectra matching Observed Spectra

6. Standards Information
  - a. Enter name of standard A
  - b. Enter name of standard B
  - c. Enter relevant standard concentrations for Standard A and Standard B
  - d. Lot Number, Purity, Std. Weight, Preparation Date - informational
7. Bath - for Distek baths only
  - a. Select Enable Bath A and or B
  - b. Set Temperature, RPM, Medium and Set Vol. (volume)
  - c. Select Active Vessel Positions

## Acquisition of Blanks and Standards

1. Standard Blanks
  - a. Place probes in dissolution media or media used in preparation of standards
  - d. Acquire Standard Blank A and or Standard Blank B by selecting each individually under the Acquire Tab
    - i. The intensity of the lowest peak must be at least 40% of the highest peak
    - ii. Select Spectral View
      1. Consistency: With all blank readings you are looking for consistency (If probe 3 is noticeably the strongest in intensity it should remain the strongest throughout all blank readings)
      2. Similar Shape: All probes, though different intensities, should have a similar shape (A strange shape is indicative of an air bubble)
2. Standards
  - a. Move probes from tank with blank media into tank with standard media blotting dry with Kimwipe (If two standards are prepared pour standards into two separate tanks and label accordingly)
  - b. Acquire Standard A and Standard B by selecting each individually under the Acquire tab
    - i. The absorbance of the individual channels may differ slightly due to the slight differences in the true pathlengths
    - ii. Select Spectral View
      1. Similar Shape: Each channel's absorbance plot should have similar shaped curves
    - iii. Select Report View
      1. Verify the Response Factor ratio of the two standards is 1.00 plus or minus 0.02
3. Sample Blanks
  - a. Non-mixture mode (and Single Component)
    - i. Place probes into individual vessels using cover spacer for 900 mL test and direct into cover for 500 mL test
    - ii. Acquire Sample Blank under Acquire tab
  - b. Mixture mode
    - i. Place probes in dissolution media or media used to prep standards
    - ii. Acquire Sample Blank under Acquire tab
4. Standard Mixtures
  - i. Place probes in tank with standard mixture
  - ii. Acquire mixture spectra as Ad Hoc datasets using Acquire Image under Acquire tab

- iii. Move probes to next mixture tank after blotting dry with Kimwipe and Acquire Image
- iv. Repeat for all mixtures
- v. Place probes in vessels either using cover spacer for 900 mL test or directly into the cover slot for 500 mL test

## Starting a Run

1. Dissolution Unit Set-up
  - a. Make sure dissolution unit is set up with correct RPM and run time if not in communication with Opt Diss
  - b. Verify vessel temperatures have reached 37.0 degrees  $\pm$  0.5
2. Preparation of Dosage Forms
  - a. Baskets
    - i. Raise Drive Head
    - ii. Place dosage form in basket
    - iii. Attach basket to adapter
  - b. Paddles
    - i. Locate dosage forms as close to hole in vessel cover as possible
    - ii. Stop rotation of the paddle blades
3. Starting the Opt Diss
  - a. Start Run
    - i. Select Start Run under Acquire Tab
    - ii. Enter appropriate drop timer
4. Starting the Test
  - a. Select OK from drop timer window
  - b. Lower drive head for baskets
  - c. Drop dosage forms for paddles
  - d. Start rotation of stirring device